antigens. This test is variously referred to as the PPD test, the tuberculin skin test, and the Mantoux test, among other designations. Unfortunately, the cutaneous immune response to these tuberculosis-derived filtrate proteins does not differentiate well between infection with some NTM and that with *M. tuberculosis*. Because intermediate reactions (~10 mm) to PPD in latent tuberculosis and nontuberculous mycobacterial infections can overlap significantly, the progressive decline in active tuberculosis in the United States means that NTM probably account for increasing proportions of PPD reactivity. In addition, *bacille Calmette-Guérin* (BCG) can cause some degree of cross-reactivity, posing problems of interpretation for patients who have received BCG vaccine. Assays to measure the elaboration of IFN-γ in response to the relatively tuberculosis-specific proteins ESAT6 and CFP10 form the basis for IFN-γ-release assays (IGRAs). These assays can be performed with whole blood or on membranes. It is important to note that *M. marinum, M. kansasii*, and *M. szulgai* also have ESAT6 and CFP10 and may cause false-positive reactions in IGRAs. Despite cross-reactivity with NTM, large PPD reactions (>15 mm) most commonly signify tuberculosis. Conversely, in the setting of anti-IFNg autoantibodies the IGRA test is indeterminate (failure of IFNg detection in response to specific antigens and mitogens, due to neutralizing anti-IFNg autoantibodies).

Isolation of NTM from blood specimens is clear evidence of disease. Whereas rapidly growing mycobacteria may proliferate in routine blood culture media, slow-growing NTM typically do not; thus it is imperative to suspect the diagnosis and to use the correct bottles for cultures. Isolation of NTM from a biopsy specimen constitutes strong evidence for infection, but cases of laboratory contamination do occur. Identification of organisms on stained sections of biopsy material confirms the authenticity of the culture. Certain NTM require lower incubation temperatures (*M. genavense*) or special additives (*M. haemophilum*) for growth. Some NTM (e.g., *M. tilburgii*) remain noncultivable but can be identified molecularly in clinical samples.

The radiographic appearance of nontuberculous mycobacterial disease in the lung depends on the underlying disease, the severity